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Toxic effects of four biopesticides (Mycotal, Vertalec, Vertemic and Neem Azal-T/S) on *Bemisia tabaci* (Gennadius) and *Aphis gossypii* (Glover) on cucumber and tomato plants in greenhouses in Egypt

ABSTRACT

Laboratory bioassays and field trials were performed to evaluate and compare the efficacy of 4 biopesticides, Mycotal, Vertalec, Vertemic and NeemAzal-T/S, against *Bemisia tabaci* (immature stages and adults) and *Aphis gossypii* on cucumber and tomato plants in greenhouses in Egypt. All these biopesticides reduced significantly whitefly and aphid populations. In laboratory bioassays Vertemic caused the highest toxic effect against whitefly and aphid, while in field experiments good results were obtained with all the compounds especially at high concentrations. The higher the concentration of these products the more severe was the effect. The data of this study suggest that these biopesticides are really useful alternative products to be used in IPM of tomato and cucumber crops.

Key words: whitefly, aphid, Verticillium lecanii, Abamectin, Azadirachtin.

INTRODUCTION

Cucumber (*Cucumis sativus* L.) and tomato (*Solanum lycopersicum* L.) plants are considered two of the most important vegetable crops in Egypt. They are attacked by many insect species including whitefly, *Bemisia tabaci*, (Gennadius, 1889) (Rhynchota: Aleyrodidae) and *Aphis gossypii* Glover, 1877 (Rhynchota: Aphididae).

The sweetpotato whitefly, *Bemisia tabaci* is an important pest on a variety of crops, causing direct damage (such as physiological disorders and reduction of plant development) and transmits more than 70 different plant viruses (BERLINGER & DAHAN, 1987; BROWN *et al.*, 1989). *B. tabaci* comprises sibling species and/or biotypes which causes great economical damage to field and greenhouse crops, reaching up to 100 % of losses (BACCI *et al.*, 2007) in subtropical and tropical regions (OLIVEIRA *et al.*, 2001). *B. tabaci* is also important within a social context for increasing levels of

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unemployment in some rural areas (BACCI *et al.*, 2007). Infestation by *B. tabaci* modifies the vegetative and reproductive development of the injured plant and causes direct damage by sucking the plant sap and injecting toxins. The latter causes physiological changes including leaf silvering in cucurbits (COSTA *et al.*, 1993) irregular ripening of tomatoes (SCHUSTE *et al.*, 1990), foliar disorders in ornamentals (TSAI *et al.*, 1997) and white streaking on *Brassica* species (BROWN *et al.*, 1992). *B. tabaci* also causes indirect damage by excretion of honeydew (BROWN *et al.*, 1995); the growth of sooty mould on honeydew decreases photosynthesis and damages harvestable parts (LIU, 2000).

In Egypt, aphids are considered among the most serious pests of different plants as well as all over the world. Many species are common as major pests attacking economic crop plants. In addition, some species are playing a great role as a vector of certain plant virus diseases (AHMED *et al.*, 2007). The cotton or melon aphid, *Aphis gossypii*, is a polyphagous species widely distributed in tropical, subtropical and temperate regions (LECLANT & DEGUINE, 1994). This aphid is a pest of cotton, cucurbits and citrus, and in temperate zones principally attacks vegetables in fields and greenhouses (BLACKMAN & EASTOP, 1985). *A. gossypii* causes direct damage, inducing pant deformation and indirect damage caused either by honeydew or by transmission of viruses (KING *et al.*, 1987; SLOSSER *et al.*, 1989; GONZALES *et al.*, 1992). The cotton aphid is the vector of 76 virus diseases in a very large range of plants (CHAN *et al.*, 1991).

In recent years, however, *A. gossypii* has emerged as one of the main aphid species on citrus, cotton, cucurbits, and greenhouse-grown vegetables (SATAR *et al.*, 1999). The cotton aphid is currently becoming more important pest because of increased insecticide tolerance and destruction of natural enemies through the use of chemical pesticides in cotton and vegetable plantations (GHABEISH *et al.*, 2008). The cotton aphid is an increasing problem in glasshouse vegetables. Presently it can only be controlled by non-selective chemicals, which inhibits the use of biological control of other pests in the glasshouse.

Synthetic chemical insecticides have played important and beneficial roles in the control of agricultural pests and the reduction of insect borne diseases for nearly 50 years. Their use will remain essential for many more years. Nonetheless, insecticides also pose real hazards. Some leave undesirable residues in food, water and environment. Low doses of many insecticides are toxic to humans and other animals, and some insecticides are suspected to be carcinogens. An effective way to delay resistance to insecticides and still maintain insect population densities below the economic threshold is to reduce the use of pesticides with the integration of other control strategies (TABASHNIK, 1986). As a result, many researchers and farmers are seeking less hazardous alternatives to conventional synthetic insecticides.

Microbial insecticides are composed of microscopic living organisms (viruses,

bacteria, fungi, protozoa or nematode-bacteria complexes) or the toxins produced by these organisms. They are formulated to be applied as conventional insecticidal sprays, dusts or granules. Each product's specific properties determine the ways in which it can be used most effectively. The fungal pathogens will be suitable for greenhouse crops (KANAGARANTNAM *et al.*, 2008; SANTIAGO-ÁLVAREZ *et al.*, 2006). The organisms already used on whitefly and aphids are likely to be commercially available. These include the two fungal pathogens, *Verticillium lecanii* (Zimmerm.) Viegas and *Paecilomyces fumosoroseus* (Wize) Brown & Smith. Other natural enemies are available overseas for growers to use.

Abamectin, as Spinosad, are microbial derived biorational pesticides used against insect pests of several orders worldwide. Abamectin acts through ingestion and contact and kills the insects through targeting the nervous system by a persistent activation of acetylcholine receptors (THOMPSON *et al.*, 2000; COWLES *et al.*, 2000; TJOSVOLD & CHANEY, 2001). Similarly, Spinosad also target insects by affecting the function of nerve fibers.

Azadirachtin (product Neem Azal-T/S), a steroid-like tetranortriterpenoid derived from neem trees, *Azadirachta indica* A. Juss. (family Meliaceae), is a strong antifeedent, repellent and growth-regulating compound for a variety of phytophagous insects, including whiteflies (COUDRIET *et al.*, 1985; SCHMUTTERER, 1990). It delays or prevents molting; reduces growth, development, and oviposition; and can cause significant mortality, particularly in immature stages (COUDRIET *et al.*, 1985; FLINT & SPARKS, 1989; PRABHAKER *et al.*, 1989; SCHMUTTERER, 1990; LIU & STANSLY, 1995; MITCHELL *et al.*, 2004).

The present study aimed to provide and evaluations of the effectiveness of 4 alternative products that are available for use in insect pest management: Mycotal, Vertalec, Vertemic and NeemAzal-T/S.

MATERIALS AND METHODS

Products

- Mycotal and Vertalec are a wettable powders based on the spores of a specific strain of *Verticillum lecanii*. The products contain 10¹⁰ spores/gram and the spray solution 10⁷ spores/ml (VAN DER PAS *et al.*, 1995). Each compound was applied at the rates 4g/lt (4X10⁷ spores/ml), 3g/lt (3X10⁷ spores/ml), 2g/lt (2X10⁷ spores/ml) and 1g/lt (1X 10⁷ spores/ml) (Table 1).

- Abamectin (Vertemic 1.8% EC) is a mixture of avermectins containing about 80% avermectin B1a and 20 % avermectin B1b; B1a and B1b have very similar biological and toxicological properties. The avermectins are insecticidal and anthelmintic

compounds derived from various laboratory broths fermented by the soil bacterium *Streptomyces avermitilis* and Abamectin is a natural fermentation product of this bacterium. No recommended dose rates for abamectin against whiteflies and aphids were available but dose rates chosen were 1–4 ml/lt (Table 1) water based on recommended rate of 1– 4 ml of abamectin for *Plutella xylostella* (L.), *Helicoverpa armigera* (Hübner) and *Spodoptera* spp. on brassicaceous crops (KUMAR & POEHLING, 2007). Vertemic was applied at rates of 6 ml/lt and 4 ml/lt water (Table 1)

- NeemAzal-T/S 1% consists of 4 % NeemAzal-T/S i.e. 1% Azadirachtin A (10g/L) and 3% other neem substances. It contains about 51% plant oil. The product is now registered in Germany as commercial insecticide for the control of greenhouse and some field pests. It was applied at the rates of 10 ml/lt and 5 ml/lt water (Table 1).

Three sprays were applied with one week intervals for all these compounds.

INSECTS

	Table 1 - Bioinsecticic	le products	s: rates of app	lication i	in lab	bioassays
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	Mycotal against B. tabaci on cucumber and tomato plants							
Concentrations	g/liter (a.i)	Dilution						
M 1	4 g	$4 \ge 10^7$						
M 2	3 g	3×10^{7}						
M 3	2 g	2×10^{7}						
M 4	1 g	1×10^{7}						
	Vertalec against A. gossypii o	on cucumber and tomato plants						
	g/liter (a.i)	Dilution						
V 1	4 g	$4 \ge 10^7$						
V 2	3 g	3×10^{7}						
V 3	2 g	2×10^{7}						
V 4	1 g	$1 \ge 10^{7}$						
	Abamectin (Vertemic) on o	amectin (Vertemic) on cucumber and tomato plants						
	g/liter (a.i)	Dilution						
A 1	0.108 ml / liter	6 ml / liter						
A 2	0.072 ml / liter	4 ml / liter						
	Neem – Azal T/S on cuc	cumber and tomato plants						
	g/liter (a.i)	Dilution						
N 1	0.05 g/liter	5 ml / liter						
N 2	0.1 g / liter	10 ml / liter						

- *Bemisia tabaci*. Infested cucumber and tomato leaves with immature stage of *B*. *tabaci* were taken from an agricultural greenhouse and transferred to the laboratory.

- Aphis gossypii. The stock culture of cotton aphid was collected from infested cotton seedling leaves in the laboratory. A. gossypii was reared in a controlled climate room at 20 ± 2 °C and 60 ± 5 % R.H. Pots of cotton seedlings cultivated in soil were placed in center of the cage (measured 80X60X60 cm). A continuous light source of (60 watt) fluorescent was provided. Large numbers of A. gossypii were placed on leaf of the cotton seedling and then left to reproduce. This strain was reared for several generations in the laboratory according to NORMAN & SUTTON (1967).

BIOASSAYS

- *Bemisia tabaci*. Laboratory bioassay experiments were carried out to evaluate the relative toxicity of Mycotal, Vertalec, Abamectin and Neem Azal-T/S against immature stages of *B. tabaci* by using a leaf-dipping method (PARK *et al.*, 2002). Emulsions of treatments were prepared by mixing a few drops of Triton-X on water. Serials of concentrations for each of the tested products were prepared. Infested plant leaves with immature stages of *B. tabaci* were dipped into the treatment solution for 10s. Twenty five leaves were used for each concentration (5 leaves per replicate). Control leaves were left for dryness, and then kept in incubator. Counts of mortality of whitefly were carried out after 2 days for treatment with Abamectin and Neem Azal T/S and after 7 days for the treatment with Mycotal and Vertalec.

- Aphis gossypii. Toxicity of the tested compound was investigated against *A. gossypii* using residue film technique as described by IWUALA *et al.* (1981). Different concentrations of the compounds by mixing a few drops of Triton-X on water. A known volume of all tested concentrations was evenly spread at the bottom of Petri-dish surface 7 cm in diameter and kept until dryness. Five concentrations for each treatment were used and each one was replicated five times. After complete film dryness, the number of adults were placed in each of the treated Petri dishes, covered and incubated at 20±2 °C, and the percentage mortality was calculated after 24 h. for the Abamectin and Neem Azal T/S and after 7 days for the Mycotal and Vertalec. Control was prepared with water and Triton-X only. Corrected mortality counts according to formula (ABBOTT, 1925), then submitted to probit analysis using FINNEY (1971):

Relative toxicity = $\frac{LC_{50} \text{ of the most active treatment}}{LC_{50} \text{ of certain treatment}} \times 100$

FIELD TRIALS

The efficiency of these compounds were tested in cucumber and tomato crops in commercial greenhouses at the farm of Faculty of Agriculture in Qena, Egypt. The greenhouse temperature ranged between 28°C to 30°C in daytime and between 20°C to 22°C at night. The experiment was carried out during November 2007–January 2008. The experimental area was divided according to complete randomized block design including 5 replicates for each treatment.

All insecticides were sprayed 3 times at weekly intervals with a knapsack sprayer. Samples of 10 leaves each were taken at random from each replicate representing different levels of the plants. They were examined in greenhouses in early morning to estimate the living adults of whitefly and then taken to laboratory to calculate the alive number of immature stages with the aid of binocular microscope. All counts of treatments and control were recorded shortly before spraying and at 3 and 7 days after each spray. Percent reduction in infestation of immature and adult stages of whitefly and apterous adults of aphid for each treatment were calculated according to HENDERSON & TILITON (1955) formula.

STATISTICAL ANALYSIS AND TOXICITY LINES

Differences between the percentages of host mortality were evaluated by analysis of variance (STATISTICA, 1999). Means were separated using Tukey's test-comparison procedure (significance level, p < 0.05).

RESULTS AND DISCUSSION

Laboratory bioassays. Toxicity of tested compounds against *B. tabaci* and *A. gossypii*. Results concerning the toxic effect of entomopathogenic fungi products, Mycotal and Vertalec, on different stages of *B. tabaci* and *A. gossypii* are shown in Fig. 1. Data clarify that the tested substances caused toxic effects on all the tested insect. Results indicate that the LC₅₀ entomopathogenic fungi products, were 7.299 gm/L and 7.936gm/L for Mycotal and Vertalec respectively.

Results concerning the toxic effect of Abamectin and Neem Azal T/S on different stages of *B. tabaci* and *A. gossypii* are shown in Fig. 2. The bioinsecticide Abamectin was the most toxic to whitefly and aphids. The LC₅₀ of the tested compounds were 2.873 ml/L, 3.152 ml/L and 10.921 and 12.50 ml/L for *B. tabaci* and *A. gossypii*, respectively.

FIELD EVALUATION

Effects of different concentrations of Mycotal and Vertalec on *B. tabaci* and *A. gossypii* on cucumber and tomato plants:

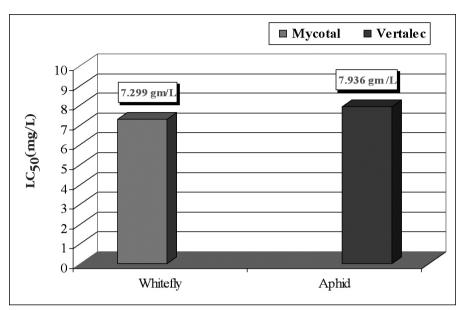


Fig. 1 - Toxicity of Mycotal against B. tabaci and Vertalec against Aphis gossypii.

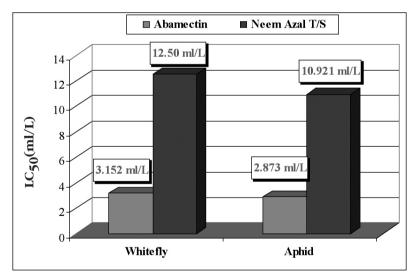


Fig. 2 - Toxicity of Vertimec and Neem Azal T/S against B. tabaci and A. gossypii.

Results are set out in Figs. 3, 4, 5, 6, 7, 8, 9 and 10 and Tables 2, 3 and 4. Data presented in Figs. 3 and 4 showed the mean mortality percentages of adult and immature stages respectively of *B. tabaci* in cucumber plants after treated with different

concentrations of Mycotal. Results indicated that there were significant differences between the control and the concentrations of Mycotal (F=187.45; df=1.39, P<0.005) and also significant differences occurred between the high (M1, M2) and low (M3, M4) concentrations and their toxic effects against whitefly stages (F=7.0; df=1.21). Data presented in Fig. 5 showed also the mean mortality percentages of *A. gossypii* in cucumber plants after treated with different concentrations of Vertalec; in this case there were no significant differences among concentrations (F=2.0; df=1.18).

Data presented in Figs. 6 and 7 showed the mean mortality percentages of *B. tabaci*, adult and immature stages respectively, in tomato plants after treated with different concentrations of Mycotal. Results indicated that there were significant differences between the control and the concentrations of Mycotal (F=222.44, df=1,15, P <0.005) and also significant differences occurred between the low and high concentrations and their toxic effects against whitefly stages (F=6.3; df=1.28). Data presented in Fig. 8 showed also, the mean mortality percentages of *A. gossypii* in tomato plants after treated with differences among concentrations of Vertalec and no significant differences among concentrations of whitefly stages and aphid in cucumber and tomato plants respectively treated three times with Mycotal and Vertalec as a comparison between 4 concentrations, after 7 days post treatment.

Data concerning percentages of reduction in infestation of immature and adult stages of whitefly in cucumber and tomato plants after 4 concentrations of Mycotal and Vertalec are set out in Tables 2, 3 and 4. The first spray was started when the mean number of 29.16 adults and 12.62 immature in cucumber plants and 37.8 adults 18.04 immature in Tomato plants, respectively. Results indicated that there were differences between the control and insecticides treatments and also, differences occurred between the different sprays and treatments. Results in Table 2 elucidate the percentage reduction in the population of *B. tabaci* during the period of three successive applications with 4 concentrations of Mycotal in cucumber plants. After the first spray M1 highly reduced the immature stages of *B. tabaci* 3 days after spraying followed by M2. Their respective percentages of reduction were 61.51 and 47.68 %. Seven days after application all concentrations of Mycotal caused more than 50 % reduction of the immature stages. Their respective percentages of reduction were 74.56, 65.15, 53.78 and 50.10 % for M1, M2, M3 and M4, respectively. The second spray was started at the seventh day of the first spray. The results showed that the four concentrations of Mycotal tested caused considerable reduction in population density of *B. tabaci* ranged between 58.27, 71.47 % and 59.28, 78.29 % at 7th days after the second spray for adult and immature stages, respectively. The alive number of adult and immature stages of whitefly decreased after seven days of third spray with the concentrations of Mycotal. It were presented 81.19, 78.74, 77.02 and 65.90 % in immature stages and 86.73, 81.86, 80.39 and 77.50 % in adult infestations by using M 1, M 2, M 3 and M 4, respectively.

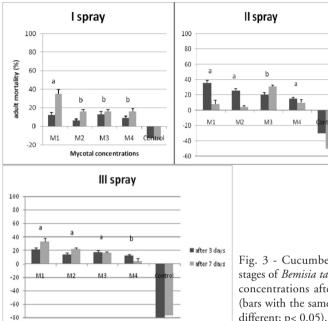


Fig. 3 - Cucumber. Mortality (%) of adult stages of *Bemisia tabaci* with Mycotal different concentrations after I, II and III treatments (bars with the same letter are not significantly different; p < 0.05).

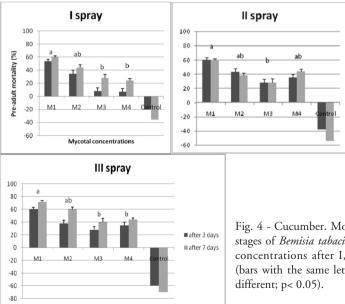


Fig. 4 - Cucumber. Mortality (%) of pre-adult stages of *Bemisia tabaci* with Mycotal different concentrations after I, II and III treatments (bars with the same letter are not significantly different; p< 0.05).

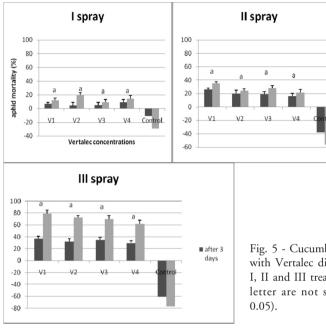


Fig. 5 - Cucumber. Mortality (%) of aphid with Vertalec different concentrations after I, II and III treatments (bars with the same letter are not significantly different; p < 0.05).

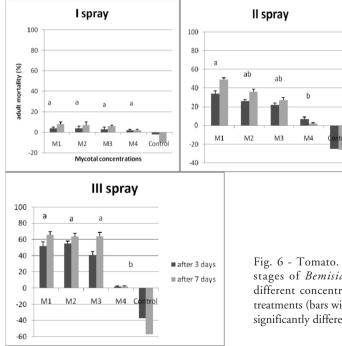


Fig. 6 - Tomato. Mortality (%) of adult stages of *Bemisia tabaci* with Mycotal different concentrations after I,II and III treatments (bars with the same letter are not significantly different; p< 0.05).

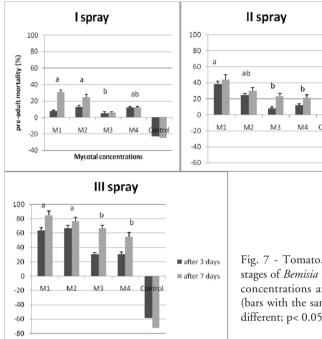


Fig. 7 - Tomato. Mortality (%) of pre-adult stages of *Bemisia tabaci* with Mycotal different concentrations after I, II and III treatments (bars with the same letter are not significantly different; p< 0.05).

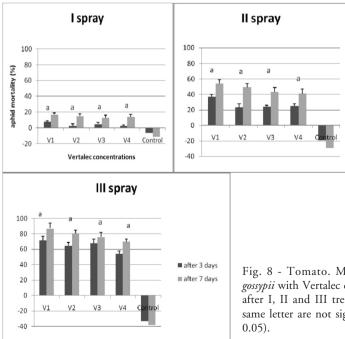


Fig. 8 - Tomato. Mortality (%) of *Aphis* gossypii with Vertalec different concentrations after I, II and III treatments (bars with the same letter are not significantly different; p< 0.05).

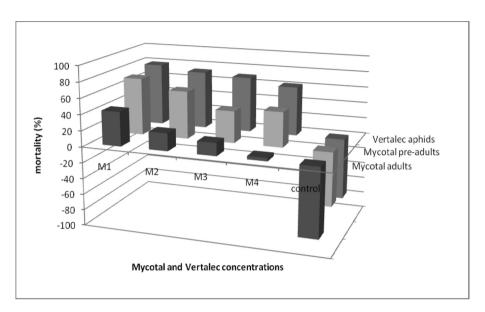


Fig. 9 - Mortality (%) of whitefly stages and aphid on cucumber plants treated 3 times with Mycotal and Vertalec as a comparison between 4 concentrations, after 7 days post treatment.

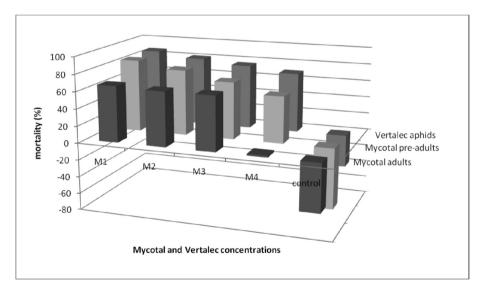


Fig. 10 - Mortality (%) of whitefly stages and aphid on tomato plants treated 3 times with Mycotal and Vertalec as a comparison between 4 concentrations, after 7 days post treatment.

Compound	N	0.	Adult stages				Pre-adult stages			
	treat	ment	М	ycotal co	ncentratio	ns	М	ycotal co	ncentratio	ons
		Period	M 1	M 2	M 3	M 4	M 1	M 2	M 3	M 4
	First	3 days	23.15	19.39	19.36	17.29	61.51	47.68	25.71	23.22
	spray	7 days	44.48	27.34	27.34	26.75	74.56	65.15	53.78	50.10
Mycotal		Mean	33.82	23.37	23.35	22.02	68.04	56.42	39.75	36.66
	Second	3 days	30.50	30.23	25.62	19.73	77.37	68.53	60.89	58.65
	spray	7 days	71.47	70.08	68.34	58.27	78.29	70.56	61.37	59.28
		Mean	50.99	50.16	46.98	39.00	77.83	69.55	61.13	58.96
	Third	3 days	73.10	67.70	55.09	53.27	80.91	72.76	67.91	60.61
	spray	7 days	86.73	81.86	80.39	77.50	81.19	78.74	77.02	65.90
		Mean	79.92	74.78	67.74	65.39	81.05	75.75	72.47	63.23
	General mean		54.90	49.44	46.02	42.14	75.64	67.24	57.78	52.95

Table 2 - The percentages reduction of B. tabaci post spraying with Mycotal on cucumber plants

Table 3 - The percentages reduction of B. tabaci post spraying with Mycotal on tomato plants

Compound	No.			Adul	t stage		Pre-adult stage			
	treat	ment	M	Mycotal concentrations			Mycotal concentrations			
		Period	M 1	M 2	M 3	M 4	M 1	M 2	M 3	M 4
	First	3 days	6.67	5.35	4.22	2.65	31.43	26.28	20.11	19.45
	spray	7 days	15.22	14.52	13.94	8.59	45.51	40.34	39.74	27.94
Mycotal		Mean	10.95	9.94	9.08	5.62	38.47	33.31	29.93	23.70
	Second	3 days	47.33	41.19	37.78	24.13	56.23	48.40	36.01	35.41
	spray	7 days	58.26	48.32	42.24	24.53	63.07	55.70	51.26	48.50
		Mean	52.97	44.76	40.01	24.33	59.65	52.05	43.64	41.96
	Third	3 days	65.30	64.61	57.04	27.62	83.60	73.27	57.30	56.46
	spray	7 days	77.89	77.35	76.40	36.28	91.44	87.75	81.01	73.30
		Mean	71.60	70.98	66.72	31.95	87.52	80.51	69.16	64.88
	General	mean	45.17	41.89	38.63	20.63	61.88	55.29	47.58	43.51

Data in Table 3 showed the percentages reduction in populations of whitefly stages after treatment with different concentrations of Mycotal in tomato plants. As regard the mean of percentage reduction in adult and immature stages of *B. tabaci* after 7 days of the third pray, it is evident that M 1, M 2, M 3 and M 4 gave considerable effects against *B. tabaci*. In term of figures, percentages of reduction of immature stages were 87.52, 80.51, 69.16 and 64.88 %, respectively. The corresponding values to adult stages were 71.60, 70.98, 66.72 and 31.95 %, respectively.

Compound	No.			Cucumber plants					
	treatme	ent	Vertalec concentrations						
		Period	V 1	V 2	V 3	V 4			
	First spray	3 days	16.07	12.80	11.05	11.00			
		7 days	37.34	30.59	28.31	22.53			
Cucumber		Mean	26.71	21.70	19.68	16.77			
plants	Second spray	3 days	45.23	40.68	37.13	35.52			
		7 days	56.19	50.53	48.08	46.38			
		Mean	50.71	45.61	42.61	40.95			
	Third spray	3 days	61.31	57.16	53.56	50.46			
		7 days	87.70	84.92	81.50	77.30			
		Mean	74.51	71.04	67.53	63.88			
	General mean		50.64	46.12	43.27	40.53			
	First spray	3 days	12.96	11.88	10.45	8.76			
		7 days	26.27	23.87	23.39	22.30			
		Mean	19.62	17.88	16.92	15.53			
Tomato	Second spray	3 days	47.09	36.16	36.02	35.39			
plants		7 days	64.26	61.09	55.60	55.10			
		Mean	55.68	48.63	45.81	45.25			
	Third spray	3 days	78.84	75.12	69.17	65.64			
		7 days	88.73	85.96	82.74	78.68			
		Mean	83.97	72.15	75.96	72.16			
	General mean		53.09	46.22	46.23	44.25			

Table 4 - The percentages reduction of *Aphis gossypii* post spraying with Vertalec product on cucumber and tomato plants

Results in Table 4 indicated that 4 concentrations of Vertalec caused moderate reduction after seven days of the second spray in cucumber plants. The percent reduction in infestation were 37.34, 30.59, 28.31 and 22.53 % for V1, V2, V3 and V4, respectively. Results indicated that all concentrations of Vertalec caused reduction of infestations in the aphis after the third spray. The corresponding values were 87.70, 84.92, 81.50 and 77.30 %, for V1, V2, V3 and V4, respectively. Data also, showed that the first concentration of Vertalec was the most effective on *A. gossypii* after seven

N	lo.		Adult	stages			Pre-adult	t stages	
treatment		Abamectin Neem Azal		Abamectin		Neem Azal			
	Period	A1	A 2	N 1	N 2	A1	A 2	N 1	N 2
First	3 days	47.96	31.58	19.36	10.71	15.13	12.34	9.91	8.46
spray	7 days	52.25	39.20	25.24	12.53	33.95	20.02	17.80	12.94
	Mean	50.11	35.39	22.30	11.62	24.54	16.18	13.86	10.70
Second	3 days	64.05	51.87	34.87	25.63	37.85	21.73	22.21	18.69
spray	7 days	72.66	58.71	43.35	30.60	48.46	29.52	30.30	20.46
	Mean	68.36	55.29	39.11	28.12	43.16	25.63	26.26	19.58
Third	3 days	83.66	73.26	62.12	49.90	66.80	54.12	60.21	27.82
spray	7 days	93.40	82.96	74.72	65.38	86.22	62.54	77.04	47.60
	Mean	88.53	78.11	68.42	57.64	76.51	59.83	68.63	37.71
General n	nean	69.00	56.26	43.28	32.46	48.07	33.88	36.25	22.66
First	3 days	50.11	37.64	26.09	14.55	29.63	13.67	11.01	10.98
spray	7 days	57.07	40.95	31.34	14.69	43.78	24.02	20.98	20.94
	Mean	53.59	39.30	28.72	14.62	36.71	18.85	16.00	15.96
Second	3 days	62.09	45.97	36.76	23.01	50.67	24.97	25.76	25.31
spray	7 days	71.73	57.34	50.19	35.26	61.65	31.44	32.44	31.08
	Mean	66.91	51.66	43.48	29.14	56.16	28.21	29.10	28.20
Third	3 days	78.85	67.14	64.41	50.90	67.08	37.94	39.91	34.30
spray	7 days	90.48	80.02	76.55	75.57	83.96	60.24	73.03	48.22
	Mean	84.87	73.58	70.48	63.24	75.52	49.10	56.47	41.26
General n	nean	68.46	54.85	47.56	35.67	56.13	32.05	33.86	28.47
	treat First spray Second spray Third spray General n First spray Second spray Third spray	PeriodFirst3 daysspray7 daysgray7 daysspray3 daysspray7 daysThird3 daysspray7 daysfirst3 daysspray7 daysfirst3 daysspray7 daysspray7 daysspray7 daysspray7 daysspray7 daysspray3 daysspray3 daysspray7 daysspray7 daysSecond3 daysspray7 daysThird3 daysspray7 days	Hereirod Aban Period A1 First 3 days 47.96 spray 7 days 52.25 Mean 50.11 Second 3 days 64.05 spray 7 days 64.05 spray 7 days 72.66 Second 3 days 68.36 spray 7 days 93.40 Second 3 days 83.66 spray 7 days 93.40 Spray 7 days 50.11 spray 7 days 50.12 spray 7 days 50.12 spray 7 days <t< td=""><td>Here A Period A1 A2 First 3 days 47.96 31.58 spray 7 days 52.25 39.20 Mean 50.11 35.39 Second 3 days 64.05 51.87 spray 7 days 72.66 58.71 spray 7 days 72.66 58.71 spray 7 days 73.26 59.21 fmird 3 days 83.66 73.26 spray 7 days 93.40 82.96 spray 7 days 50.11 37.64 spray 3 days 50.11 37.64 spray 7 days 57.07 40.95 spray 3 days 50.11 57.34 spray 7 days 71.73 57.34 <td< td=""><td>Introduction of the section of</td><td>Intraction of transmission of transmissic transmissi definitity of transmission of transmission of transmi</td><td>Intraction of the section of t</td><td>Image: Figure 1 Necetion Necetion Aba\rightarrowtrin Necetion Aba\rightarrowtrin Necetion Period A1 A2 N1 N2 A1 A2 First 3 days 47.96 31.58 19.36 10.71 15.13 12.34 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.10 34.87 25.63 37.85 21.73 Mean 68.36 75.21 31.64 25.63 69.00 <th< td=""><td>Intrave for the section of the section in the</td></th<></td></td<></td></t<>	Here A Period A1 A2 First 3 days 47.96 31.58 spray 7 days 52.25 39.20 Mean 50.11 35.39 Second 3 days 64.05 51.87 spray 7 days 72.66 58.71 spray 7 days 72.66 58.71 spray 7 days 73.26 59.21 fmird 3 days 83.66 73.26 spray 7 days 93.40 82.96 spray 7 days 50.11 37.64 spray 3 days 50.11 37.64 spray 7 days 57.07 40.95 spray 3 days 50.11 57.34 spray 7 days 71.73 57.34 <td< td=""><td>Introduction of the section of</td><td>Intraction of transmission of transmissic transmissi definitity of transmission of transmission of transmi</td><td>Intraction of the section of t</td><td>Image: Figure 1 Necetion Necetion Aba\rightarrowtrin Necetion Aba\rightarrowtrin Necetion Period A1 A2 N1 N2 A1 A2 First 3 days 47.96 31.58 19.36 10.71 15.13 12.34 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.10 34.87 25.63 37.85 21.73 Mean 68.36 75.21 31.64 25.63 69.00 <th< td=""><td>Intrave for the section of the section in the</td></th<></td></td<>	Introduction of the section of	Intraction of transmission of transmissic transmissi definitity of transmission of transmission of transmi	Intraction of the section of t	Image: Figure 1 Necetion Necetion Aba \rightarrow trin Necetion Aba \rightarrow trin Necetion Period A1 A2 N1 N2 A1 A2 First 3 days 47.96 31.58 19.36 10.71 15.13 12.34 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.87 34.87 25.63 37.85 21.73 Second 3 days 64.05 51.10 34.87 25.63 37.85 21.73 Mean 68.36 75.21 31.64 25.63 69.00 <th< td=""><td>Intrave for the section of the section in the</td></th<>	Intrave for the section of the section in the

Table 5 - The percentages reduction of whitefly insect, *B. tabaci* stages post spraying with Vertemic and Neem Azal T/S product on Cucumber and Tomato plants.

days of application in tomato plants which gave 87.70 % reduction in the aphid populations. Regarding to the mean percent reduction in infestations for the Vertalec concentrations, V1, V2, V3 and V4 were 88.73, 85.96, 82.74 and 78.68 %, respectively.

Effects of different concentrations of Vertimec and Neem Azal T/S on *B. tabaci* and *A. gossypii*:

Mortality rates of adult and pre-adult stages of *B. tabaci* and *A. gossypii* on cucumber and tomato plants at the various concentrations are shown in Figs. 11-18. There were statistically differences in the mortalities of *B. tabaci* stages caused by the 2 bioinsecticides (F=7.3, df=1.21, P<0.005), concentrations (F=6.9, df=1.12) and the two-way interaction (insecticides-concentrations) (F=5.4, df=1.21). Abamectin increased the mortality of *B. tabaci* populations compared with Neem Azal T/S on

	No. Spra	No. Sprays		Vertemic		n Azal
		Period	Al	A 2	N 1	N 2
	First	3 days	18.76	15.52	20.70	20.52
	spray	7 days	30.62	24.30	28.27	25.87
Cucumber		Mean	24.69	19.91	24.49	23.20
	Second	3 days	41.27	32.48	36.27	32.47
	spray	7 days	47.24	36.52	39.23	33.64
		Mean	44.26	34.50	37.75	30.06
	Third	3 days	57.05	40.70	42.40	34.02
	spray	7 days	81.10	46.28	47.70	36.76
		Mean	69.08	43.49	45.05	35.39
	General	mean	46.01	32.63	35.76	29.55
	First	3 days	18.71	11.30	18.26	11.30
	spray	7 days	35.46	27.00	30.20	25.07
		Mean	27.09	19.15	24.23	18.19
	Second	3 days	44.71	30.91	31.40	30.30
Tomato	spray	7 days	49.46	35.85	36.67	33.76
		Mean	47.09	33.38	34.04	32.03
	Third	3 days	65.93	40.42	60.59	38.93
	spray	7 days	89.78	70.19	71.00	40.53
		Mean	77.86	55.31	65.80	39.73
	General	mean	50.68	35.95	41.36	29.98

Table 6 - The percentages reduction of *Aphis gossypii* post spraying with Vertemic and Neem Azal T/S products on cucumber and tomato plants.

cucumber plants (Table 5). Results indicate that 93.40 and 82.96 %, 74.72 and 65.38 % as initial kill of *B. tabaci* adult and pre-adult stages and 86.22 and 62.54 % and 77.04 and 47.60 % for two concentrations of Abamectin (A1, A 2) and (N 1, N 2) after 7 days of the third spray on cucumber plants, respectively. As regards, the mean of reduction percent in whitefly, *B. tabaci* pre-adult and adult stages in tomato plants after the 7 days of the third spray it is evident that the first concentration of Abamectin (A1) was the most effective on the adult and pre-adult stages of *B. tabaci* causing 90.48 and 83.96 % reduction, respectively (Table 5).

The results presented in Table 6 indicated the effects of Vertemic and Neem Azal T/S concentrations on *A. gossypii* in cucumber and tomato plants. For tomato the

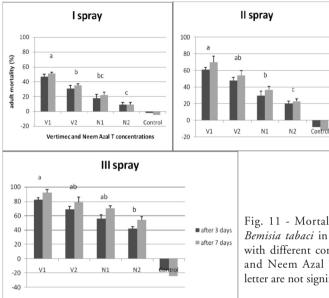


Fig. 11 - Mortality (%) of adult stage of *Bemisia tabaci* in Cucumber plants treated with different concentrations of Abamectin and Neem Azal T/S (bars with the same letter are not significantly different; p< 0.05).

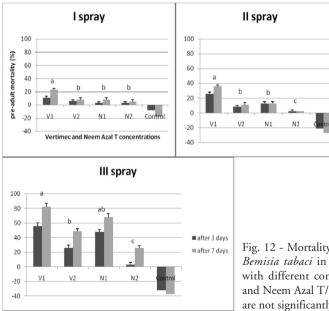


Fig. 12 - Mortality (%) of pre-adult stages of *Bemisia tabaci* in Cucumber plants treated with different concentrations of Abamectin and Neem Azal T/S (bars with the same letter are not significantly different; p< 0.05).

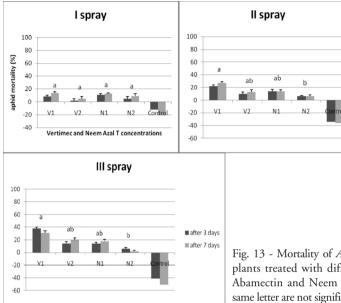


Fig. 13 - Mortality of *Aphis gossypii* in cucumber plants treated with different concentrations of Abamectin and Neem Azal T/S (bars with the same letter are not significantly different; p< 0.05).

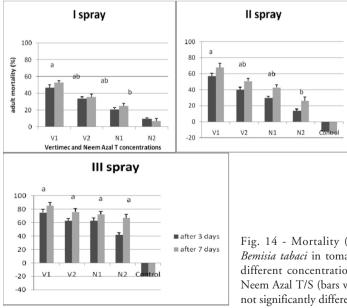
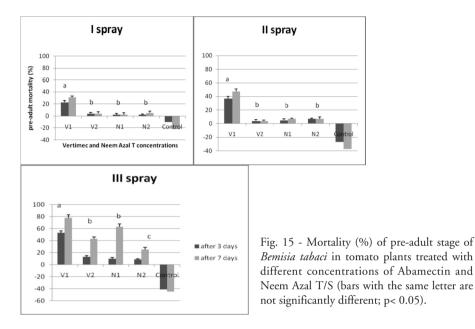


Fig. 14 - Mortality (%) of adult stage of *Bemisia tabaci* in tomato plants treated with different concentrations of Abamectin and Neem Azal T/S (bars with the same letter are not significantly different; p< 0.05).



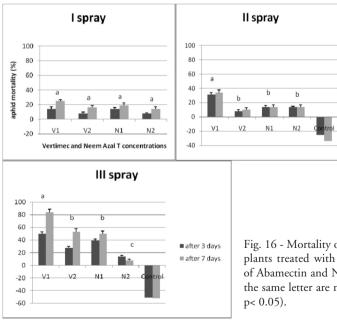


Fig. 16 - Mortality of *Aphis gossypii* in tomato plants treated with different concentrations of Abamectin and Neem Azal T/S (bars with the same letter are not significantly different; p < 0.05).

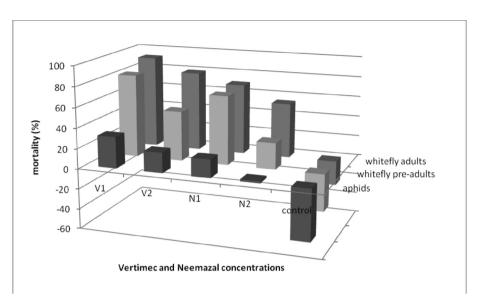


Fig. 17 - Mortality (%) of whitefly stages and aphid in cucumber plants treated 3 times with Abamectin and Neem Azal T/S as a comparison between 2 concentrations for each, after 7 days post treatment.

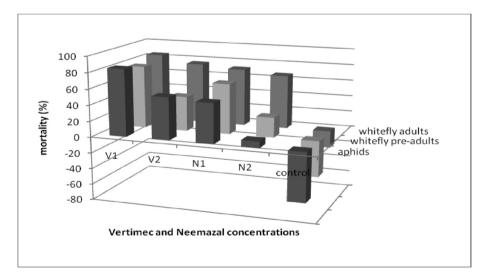


Fig. 18 - Mortality (%) of whitefly stages and aphid in tomato plants treated three times with Abamectin and Neem Azal T/S as a comparison between 2 concentrations for each, after 7 days post treatment.

highest percent reductions in infestations were 89.78, 70.19, 71.00 and 40.53% for Abamectin (A1, A2) and Neem Azal T/S (N1, N2), respectively; for cucumber the maximum values of mortality (%) were 81.10, 46.28, 47.70 and 36.76 % for Abamectin (A1, A2) and Neem Azal T/S (N1, N2), respectively.

In conclusion the laboratory bioassays and field trials carried out in this research have shown the efficacy of Mycotal, Vertalec, Vertemic and NeemAzal-T/S9, against *Bemisia tabaci* (immature stages and adults) and *Aphis gossypii* on cucumber and tomato plants inside greenhouses in Egypt. All these biopesticides reduced significantly whitefly and aphid populations and their use may contribute to the development of sustainable production systems through a reduction in the use of chemical insecticides and, consequently, a reduction of chemical residues on produce and insecticide resistance.

RIASSUNTO

Tossicità di quattro bioinsetticidi (Mycotal, Vertalec, Vertemic e Neem Azal-T/S) su *Bemisia tabaci* (Gennadius) e *Aphis gossypii* (Glover) in colture protette di zucchino e pomodoro in Egitto

Effetti tossici di 4 biopesticidi, Mycotal, Vertalec, Vertemic e Neem Azal–T/S su *Bemisia tabaci* (Gennadius) e *Aphis gossypii* (Glover) in coltivazioni sotto serra di cetriolo e pomodoro in Egitto. In questo studio sono stati effettuati esperimenti di laboratorio e di campo per valutare l'efficacia di 4 bioinsetticidi, Mycotal, Vertalec, Vertemic e NeemAzal-T/S9, nei confronti di *Bemisia tabaci* (stadi giovanili e adulti) e *Aphis gossypii* su coltivazioni in serra di cetriolo e pomodoro in Egitto. Tutti i prodotti testati hanno avuto un effetto tossico su *B. tabaci* e *Aphis gossypii* riducendone le popolazioni in maniera significativa. Nei biosaggi di laboratorio il Vertimec ha evidenziato l'effetto tossico maggiore, mentre negli esperimenti di campo sono stati ottenuti buoni risultati (% di mortalità) da tutti i prodotti, soprattutto quando utilizzati alle dosi più elevate. Più alte le concentrazioni delle sospensioni bioinsetticide, maggiore l'effetto tossico ottenuto. Questi dati mostrano l'efficacia di questi prodotti e confermano il loro potenziale utilizzo nelle strategie di controllo integrato delle coltivazione di cetriolo e pomodoro in serra.

Parole chiave: aleirodidi, afidi, Verticillium lecanii, Abamectina, Azadiractina.

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